Hyperspectral Imaging for Predicting Soluble Solid Content of Starfruit

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Graphical abstract

Abstract
Hyperspectral imaging technology is a powerful tool for non-destructive quality assessment of fruits. The objective of this research was to develop novel calibration model based on hyperspectral imaging to estimate soluble solid content (SSC) of starfruits. A hyperspectral imaging system, which consists of a near infrared camera, a spectrograph V10, a halogen lighting and a conveyor belt system, was used in this study to acquire hyperspectral images of the samples in visible and near infrared (500-1000 nm) regions. Partial least square (PLS) was used to build the model and to find the optimal wavelength. Two different masks were applied for obtaining the spectral data. The optimal wavelengths were evaluated using multi linear regression (MLR). The coefficient of determination (R²) for validation using the model with first mask (M1) and second mask (M2) were 0.82 and 0.80, respectively.

Keywords: Hyperspectral Imaging; Partial Least Square Regression; Starfruit

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1.0 INTRODUCTION

Starfruit is one of Malaysia’s main exported fruits. Since 1989, Malaysia has been the largest exporter of starfruit in the world. In 2008, 3648.9 metric tons of starfruit were exported to various countries in Europe (the Netherlands, France, Germany), Middle East (Saudi Arabia, Iran, Bahrain, Turkey), and Asia (Singapore, Hongkong, Indonesia). Until June 2009, the export record shows that a total of 901.509 metric tons of starfruits had been exported [1].

In Malaysia, numerous studies have been carried out to enhance the postharvest handling of starfruit to increase the consumer acceptance and satisfaction. In spectroscopy area, recently, Omar et al. [2][3] conducted a study on nondestructive intrinsic quality measurement of starfruit such as pH, firmness dan soluble solid content of starfruit using spectroscopy method. In image processing field, Abdullah et al.[4] had developed an automated quality inspection of starfruit based on the colour of starfruit. Mokji [5] had developed an algorithm with 2-dimensional colour maps to classify starfruits into six maturity grades. Meanwhile, Amirulah et al. [1] implemented Mokji’s algorithm on Field Programmable Gates Array (FPGA). Other researchers [4][6] have also studied defect detection of starfruit.

In recent years, hyperspectral imaging (also called imaging spectroscopy) has become a popular tool for quality and safety inspection of food and agricultural products [7]. Hyperspectral imaging combines spectroscopy and imaging techniques to obtain both spatial and spectral information of an object. The hyperspectral technique can be used for inspecting defects in fruit, predicting constituents of fruit to estimate its quality, food quality evaluation and sorting fruits. In addition, this technique can be applied to measure some internal attributes of fruit simultaneously and non-destructively [8].

A number of studies have been reported on the application of hyperspectral imaging technique to evaluate quality of various fruits. This technique has been successfully used for detecting defects in various fruits such as apple [9], citrus [10], and cucumber [11][12]. For determining quality attribute measurement of fruits such as soluble solid content (SSC) and firmness, researchers have conducted studies on a number of fruits such as apple [13][14][15], strawberry [16], blueberry [17], banana [18], grapes [19], pickles [11], and tart cherries [20]. These studies have shown the potential of hyperspectral imaging for fruit quality inspection using spatial dan spectral information. However, due to its non-circular shape, no study has been reported on using hyperspectral imaging for starfruit quality inspection. Hence, in this work, two types of region of interest (ROI) are introduced for the inspection.

The main objective of this study is to investigate the potential of hyperspectral imaging for estimating the soluble solid content of starfruit. The specific objectives are (1) to use hyperspectral imaging system to acquire the hyperspectral image
of starfruit in region between 500 and 1000 nm, (2) to identify several significant wavelengths that can be utilized for predicting soluble solid content of starfruit, (3) to evaluate the use of different area measurement (region of interest) to obtain the spectral data, and (4) to develop a novel calibration model using the optimal wavelengths.

2.0 MATERIALS AND METHODS

2.1 Starfruit Samples

A total of 72 samples were obtained from a starfruit farm. The samples were selected in such a way that each grade of samples could be represented by the same number. This study applied maturity standard set by FAMA (Federal Agricultural and Marketing Authority), Malaysia, with six indices. Lower index (index 1 and 2) would indicate that the starfruit is verdant while higher index (index 5 and 6) would indicate mature fruit. The samples were scanned in hyperspectral imaging system using image acquisition for determining the soluble solid content (SSC) of samples.

2.2 Hyperspectral Imaging System

A hyperspectral imaging system (Figure 1) developed in the computer vision, video and image processing (CVVIP) laboratory, Faculty of Electrical Engineering, Universiti Teknologi Malaysia, was used for this experiment. The imaging unit consisted of a 12-bit digital NIR camera and a spectograph. A MVI 1312 Photonfocus camera was used. A V10 (Spectral Imaging Ltd, Oulu, Finland) spectograph was attached to the camera to acquire hyperspectral images. An illumination unit, which consisted of two 150 W halogen lights powered by a DC voltage regulated power supply (Line light QDF5072 and DC950H, Dollan Jenner Industries, Inc, USA) was positioned at angle of 45° to illuminate the camera’s field of view. A conveyor unit with adjustable speed was used to move and present the starfruit for imaging. Furthermore, a computer with a frame graber (National Instument) was used for acquiring and processing the hyperspectral images. MATLAB software was used to create a program for image acquisition and analysis. The size of hyperspectral image generated by this system was 460X360 pixels with 180 spectral bands (wavelengths) from 500 to 1000 nm.

Using the first method (M1), image at 733.2 nm (Figure 2a) was selected for building the mask. The image gave the maximum contrast between the starfruit and the background. Therefore, the starfruit and the background could be segmented easily by simple thresholding. Also, the image at that wavelength was smoothened using an averaging filter to remove the noise. Then, the smoothened image was segmented by using Otsu’s method [21]. Finally, the morphological operations, such as dilation and erosion, were applied to improve the quality of the image. The mask generated by this method (M1) is shown in Figure 2b.

Mask M1 was then used for creating mask M2. Centre of mass (centroid) and bounding box value of white area in Figure 3a was computed using Matlab. Accordingly, two rectangles, r1, and r2, were build. The size of the rectangles were determined such that the position of the rectangles must be in the middle on the left and right side of starfruit. Figure 3b shows mask for M2. Mask M1 and M2 were used for obtaining the spectral data of each sample. Then, the performance of each the masks was evaluated.

2.3 Collecting Spectral Data

Due to the complex shape and uneven surface of starfruit, two approaches were used for collecting the spectral data of each starfruit. The first method, M1, was used to utilize the whole starfruit area as measurement area or region of interest (ROI). The second method, M2 was used to designate only some portion of starfruit area as ROI. The binary masks were created using both methods. Then, the masks were used for calculating the relative reflectance average of ROI of each starfruit in wavelength range between 500 and 1000 nm.

![Figure 1 Hyperspectral imaging system](image1)

![Figure 2](image2)

![Figure 3](image3)
2.4 Reference Measurement

After acquiring the spectral images, measurement of soluble solid content (SSC) of samples was carried out destructively. The samples were crushed using a juice extractor. Then, the juice of each sample was measured for its soluble solid content (SSC) using a digital hand-held pocket refractometer (PAL-α, Atago Co., Ltd., Japan). The SSC values were then used as references for developing the calibration model.

2.5 Building The Calibration Model

Partial least square regression (PLSR) was used to build the calibration or prediction model. This regression is a technique that generalizes and combines all features from principal component analysis and multiple regression. Its aim is to predict or analyze a set of dependent variables from a set of independent variables or predictors. In this study, PLSR was developed to find the mathematical relationship between the spectra response and one attribute of starfruit (soluble solid content).

The predicted value of the attribute of interest $\hat{Y}$ was determined as follows[16]

$$\hat{Y} = XW_{\alpha}\beta = T_{\alpha}\beta$$  \hspace{1cm} (1)

$$W^* = (W(P'W)^{-1})$$  \hspace{1cm} (2)

where $\alpha$ is the number of PLS components, $P'$ is the wavelength loadings and $\beta$ is the regression coefficient.

The number of PLS components is determined by the complexity of the model. The complexity increases with the increase of the number of PLS components. In this study, the optimum complexity of the model was estimated by using k-fold cross validation. The sample set was randomly split into k segments. One segment was left out as a validation set. The others (k-1 segments) were used as a training set. This procedure was repeated k times so that each segment would all be used as validation set in turn. In this experiment, k was set to 10. The result was a residual matrix. The mean squared error for validation (MSE$_{CV}$) was calculated from the residual matrix.

$$MSE_{CV} = \frac{1}{n} \sum_{i=1}^{n} (y_i - \hat{y}_{iCV})^2$$  \hspace{1cm} (3)

where $y_i$ and $\hat{y}_{iCV}$ are the measured and predicted values of attribute, while n is the number of sample.

The local minimum approach [22] of MSE$_{CV}$ was used for determining the optimum number of PLS component. This approach is used to avoid overfitting. Afterwards, feature (variable) selection was done by using PLS-BETA method [23][24]. The method applies regression coefficient ($\beta$) to identify the most influential wavelength. The wavelength that has the highest absolute value was considered as the optimal wavelength. Otherwise, the wavelength that has the lowest is neglected because it would only contribute little to the model. Only selected optimal wavelengths were used to build the multi linear regression (MLR) models instead of using the whole wavelengths. The formula of MLR is as follows [18]

$$\hat{Y} = a_0 + \sum_{i=1}^{W} a_i R_{iN}$$  \hspace{1cm} (4)

where $\hat{Y}$ is the predicted value of the attribute; N is the number of optimal component; $a_0$ and $a_i$ are the regression coefficients and $R_{iN}$ is the reflectance intensity at a wavelength.

The performance of the calibration model was evaluated using MSE and the coefficient of determination ($R^2$), expressing the proportion of variance explained by the model. The formula is defined as follows[17]

$$R^2 = 1 - \frac{RSS}{TSS}$$  \hspace{1cm} (5)

where $TSS$ (Total sum of squares) = $\sum_{i=1}^{n} (y_i - \bar{y})^2$, $RSS$ (Residual sum of squares) = $\sum_{i=1}^{n} (y_i - \hat{y}_i)^2$, $\hat{y}_i$ is the predicted value, while $y_i$ is measurement value.

3.0 RESULTS AND DISCUSSION

3.1 Reflectance Spectra

Figure 4 and 5 show the reflectance spectra of three starfruits with different SSC values (°Brix) using mask M1 and M2, respectively. The curve using mask M1 had the same shape as M2 in whole wavelength range. However, both curves had different reflectance value, especially in range between 708 nm and 1002 nm.

The reflectance value of the samples increased when the SSC value was increased in wavelength between 590.4 nm and 1002 nm for M1, and between 621.2 nm and 708 nm for M2. Both figures show that reflectance value had significant change at 682.8 nm for M1 and M2. At this wavelength, a strong absorbance was observed clearly, which was due to the change of the chlorophyll pigment. Lower SSC value had stronger absorbance, meaning that the chlorophyll pigment of starfruit was high. As a result, the reflectance value decreased. In contrast, the high reflectance value or high SSC value had lower absorbance and chlorophyll pigment. The high reflectance showed high maturity grade of starfruit. This wavelength (682.8 nm), which had sensitivity to chlorophyll change, was similar to the wavelength as reported by previous researchers [16][18].

![Figure 4](image1)

Figure 4 Reflectance spectra of three samples with different SSC values using mask M1

![Figure 5](image2)

Figure 5 Reflectance spectra of three samples with different SSC values using mask M2

3.2 Optimal Wavelength Selection

Two PLS calibration models were build using the whole spectral range consisting of 180 wavelengths. The first model was established using mask M1, while the second model was created using mask M2 as ROI. To find the optimal wavelength for each model, the same procedure was applied to both models. Before optimal wavelengths selection was done, the optimum number for each of the model was determined using the local minimum approach based on MSE value. Figure 6 and 7 show the plot of MSE versus PLS components for M1 model and M2 model,
respectively. Although using different ROIs, both models had a similar local minimum at n = 5.

![MSE plot of the PLS model using mask M1](image1.png)

**Figure 6** MSE plot of the PLS model using M1

![MSE plot of the PLS model using mask M2](image2.png)

**Figure 7** MSE plot of the PLS model using M2

Figure 8 and 9 show the regression coefficient (β) plot versus the wavelength of both models using the optimal component (n = 5). The PLS-BETA method was used to find the significant wavelengths. By using Matlab, the maxima and minima of β values for both models were selected. The optimal wavelengths for M1 model were 565.2, 657.6, 702.4, 741.6, 859.2 and 943.2 nm (six wavelengths), respectively. Meanwhile, the optimal wavelengths for M2 model were 565.2, 677.2, 736, 873.2 and 943.2 nm (five wavelengths), respectively. The results showed that M2 model had lesser amount of wavelengths than M1 model despite having the same number of PLS components.

![Regression coefficient plot of M1 model with n=5](image3.png)

**Figure 8** Regression coefficient plot of M1 model with n=5

![Regression coefficient plot of M2 model with n=5](image4.png)

**Figure 9** Regression coefficient plot of M2 model with n=5

3.3 The Performance Of The Model Using The Optimal Wavelengths

The optimal wavelengths, which were selected using PLS, were evaluated using multi linear regression (MLR) for predicting the SSC value of starfruit. Table 1 shows the performance of the models built by using MLR and the optimal wavelengths. The performance was evaluated via cross validation (CV). Figure 10 and 11 show the plot of the predicted and measured SSC values for calibration and validation. In general, the model created using mask M1 had better performance than the model created using M2, although the performance of both models had small differences. For MSE value, the M1 model was lower than the M2 model for calibration and validation. Meanwhile the R² of M1 model was higher than that of the M2 model. However, M2 model had smaller number of wavelengths than M1. Both models have shown good performance because R² values are high (0.82 and 0.80 out of scale of 1). The following equations were derived based (2);

\[ \hat{y} = 7.06 - 4.63R_{565.2} - 3.43R_{657.6} + 17.62R_{702.4} - 23.51R_{741.6} + 12.86R_{859.2} + 3.81R_{943.2} \]  \hspace{1cm} (6)

\[ \hat{y} = 7.40 - 0.77R_{565.2} + 9.01R_{677.2} - 17.35R_{736} + 12.722R_{873.2} - 2.07R_{943.2} \]  \hspace{1cm} (7)

where \( \hat{y} \) is the predicted SSC value; \( R_\lambda \) is the reflectance intensity at \( \lambda \). Equation (6) was used for estimating the SSC value using mask M1. Meanwhile, (7) was used for estimating the SSC value using mask M2. These equations can be used for multispectral application. For simpler application, the model with mask M2 and five wavelengths could be considered for predicting the SSC value of starfruit non-destructively.

<table>
<thead>
<tr>
<th>ROI</th>
<th>Wavelength (nm)</th>
<th>MSE_Cal</th>
<th>MSE_Val</th>
<th>R²_Cal</th>
<th>R²_Val</th>
</tr>
</thead>
<tbody>
<tr>
<td>M1</td>
<td>565.2, 657.6, 702.4, 741.6, 859.2 and 943.2</td>
<td>0.228</td>
<td>0.274</td>
<td>0.85</td>
<td>0.82</td>
</tr>
<tr>
<td>M2</td>
<td>565.2, 677.2, 736, 873.2 and 943.2</td>
<td>0.259</td>
<td>0.305</td>
<td>0.83</td>
<td>0.80</td>
</tr>
</tbody>
</table>

![The predicted and measured SSC values of the model created using MLR, mask M1 and optimal wavelengths](image5.png)

**Figure 10** The predicted and measured SSC values of the model created using MLR, mask M1 and optimal wavelengths
4.0 CONCLUSION

In this study, a hyperspectral imaging system was used to acquire hyperspectral images of starfruit. This system was successfully applied to predict the soluble solid content (SSC) of starfruit. The models were built using different ROI, and the optimal wavelengths were evaluated. The model created using mask M1 showed better performance. For simpler application, the model created using mask M2 could be considered for estimating the SSC value of starfruit.

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